

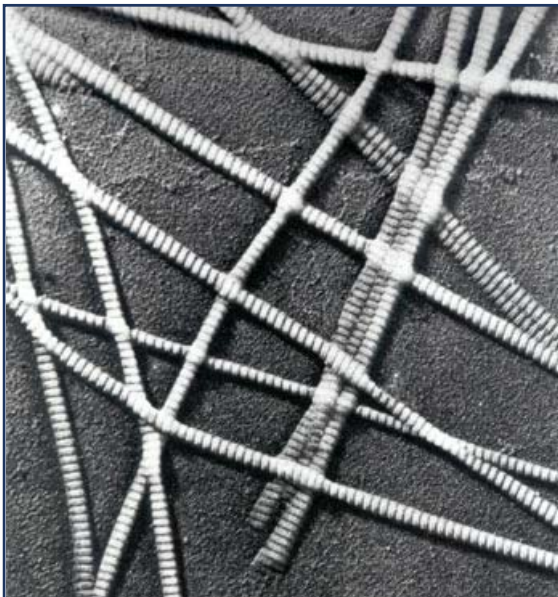
Biosensors, biocompatibilization of materials, and surface science



require the investigation of

the **sorption processes** and
the **3-D organization**
of proteins at solid/liquid interface.

Many **qualitative** methods
but
few **quantitative** methods
and
even fewer **real-time** measurements !



<http://www.aad.org/education/CollagenFigure1.htm>

Collagen

Fibrillar protein of the extracellular matrix

Triple helix 300 nm long

1.5 nm diameter

Weight ~ 300 kDa

Forms elastic fibers



Fibrinogen

Blood protein

Three globular domains linked by fibrillar segments

Weight ~ 340 kDa

Causes blood clotting

How to investigate in liquids the behavior of adsorbed proteins ?

A novel biosensor platform integrating **Love mode acoustic wave and surface plasmon resonance** for the investigation of protein layers

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OUTLINE

- Introduction

- Love mode surface acoustic wave (SAW)
 - Structure and characteristics
 - Sensitivity enhancement

- Surface plasmon resonance (SPR)

- Combined SAW/SPR
 - Technique
 - Method
 - Application to collagen and fibrinogen

- Conclusions

SURFACE ACOUSTIC WAVE

SAW

SURFACE PLASMON RESONANCE

SPR

Label-free sensing on a common surface

increased system integration

fast analysis

low analyte volume

identical event

simplicity of sample preparation

Real time measurement

adsorption kinetic

dynamic evolution of the layer

LOVE MODE SAW SENSOR: STRUCTURE

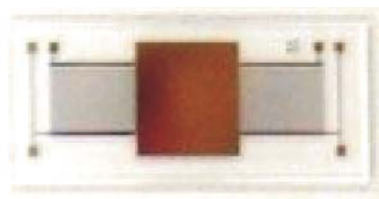
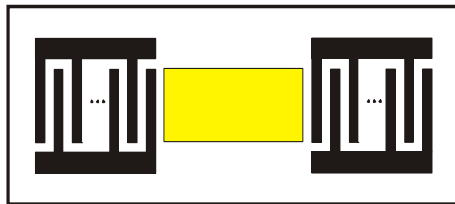
Overcoated SAW delay line is a mass sensitive device.

Shear horizontal polarized acoustic wave that results in low loss with liquid loading.

Substrates: α -SiO₂, LiNbO₃, LiTaO₃

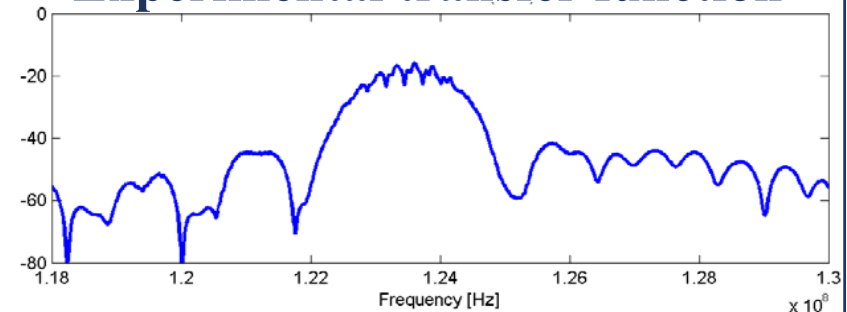
Guiding layers: SiO₂, polymers/epoxies, ZnO

Delay line structure



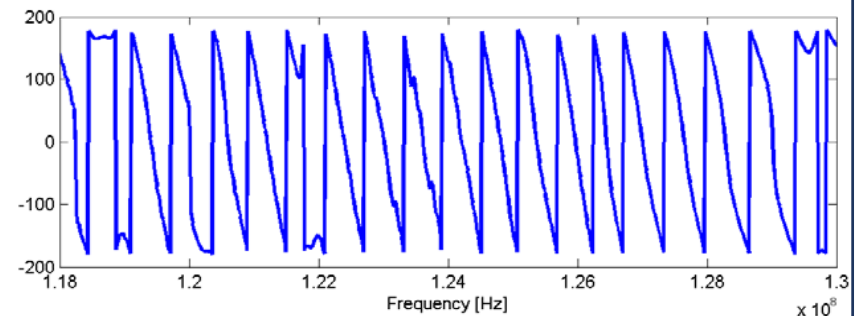
Experimental transfer function

I.L.



Phase

ϕ



LOVE MODE SAW SENSOR: CHARACTERISTICS

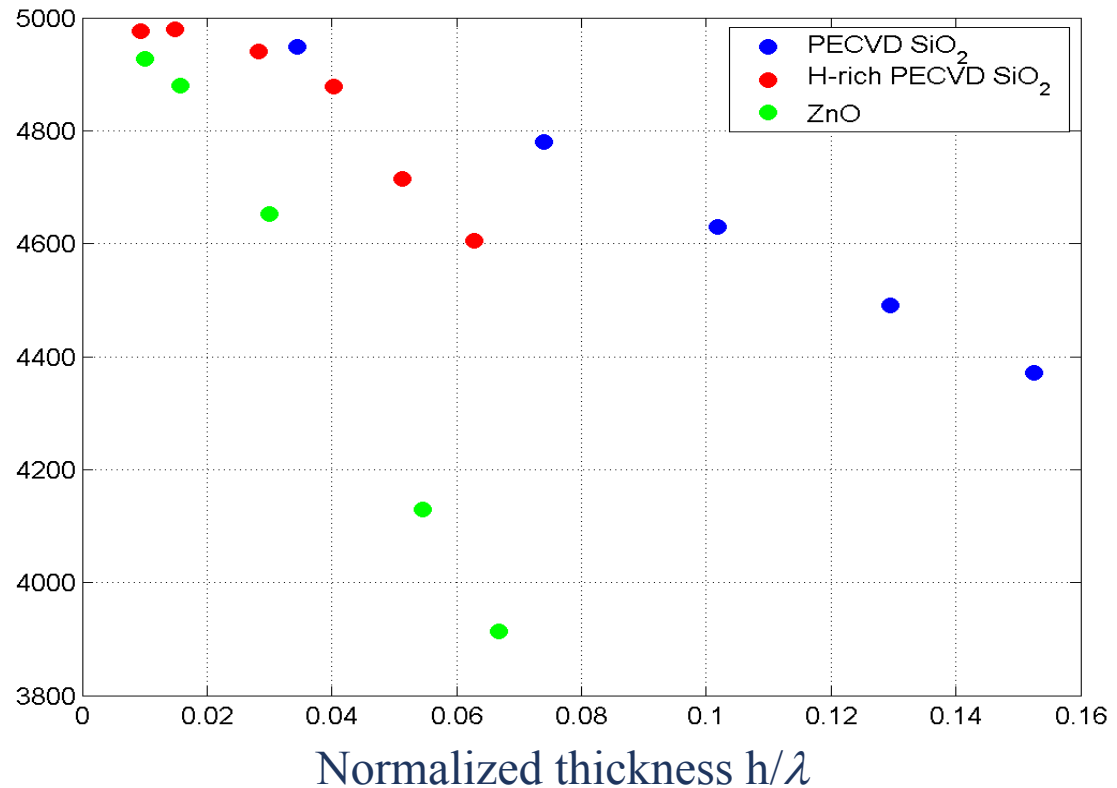
The dispersion of Love mode is ruled by

- the acoustic velocity $V^2 = \mu / \rho$
- the acoustic impedance $Z^2 = \rho * \mu$

Mechanical properties are function of

- the selected guiding materials;
- the structure (processing parameters, *i.e.* PECVD).

Experimental
phase velocity
(m/s)



LOVE MODE SAW SENSOR: MASS SENSITIVITY

Mass sensitivity

$$S = \frac{\lambda}{360^\circ D} \cdot \frac{\Delta\phi}{m}$$

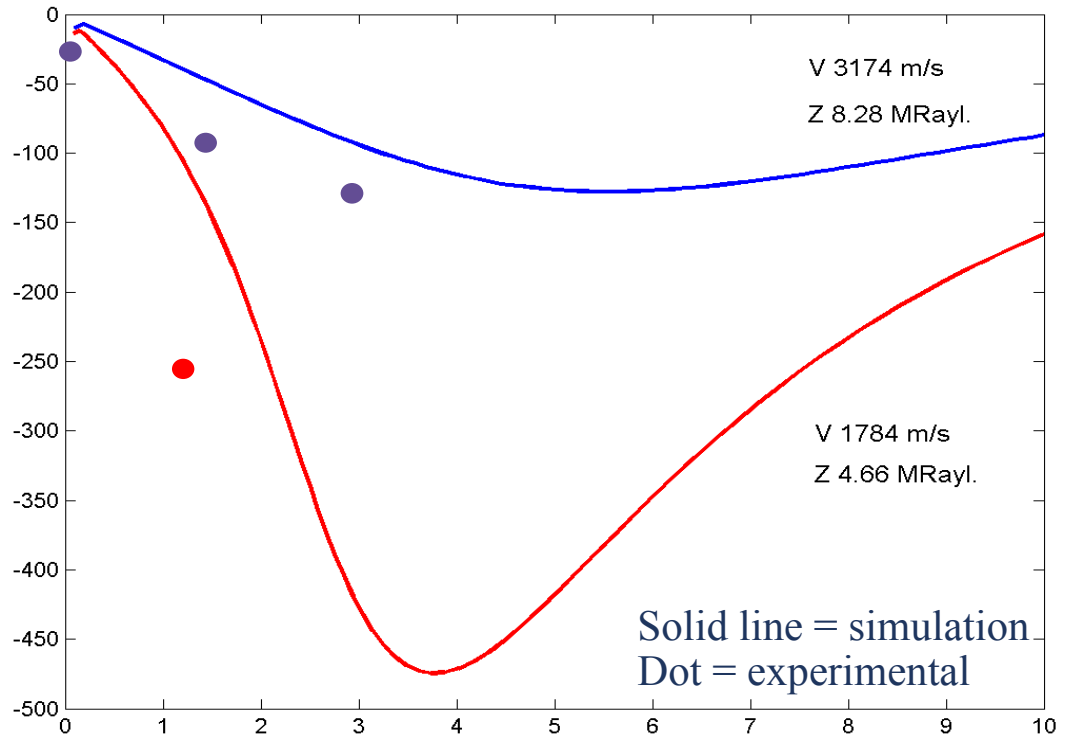
λ acoustic wavelength
 D sensing length
 m surface density

Strong dispersion = high sensitivity:

- H-rich PECVD SiO₂ (reduced stiffness);
- Gold (low velocity).

Gold (50 nm)
PECVD SiO ₂
ST-cut Quartz

S (cm²/g)



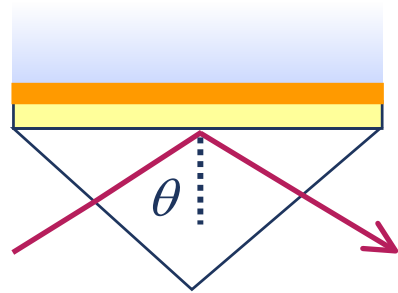
Thickness SiO₂ (µm) with $\lambda = 40$ µm

SURFACE PLASMON RESONANCE SENSOR

Surface Plasmon Resonance (SPR) is an **optical method** where collective electromagnetic modes are excited by a light source at metal-dielectric interface.

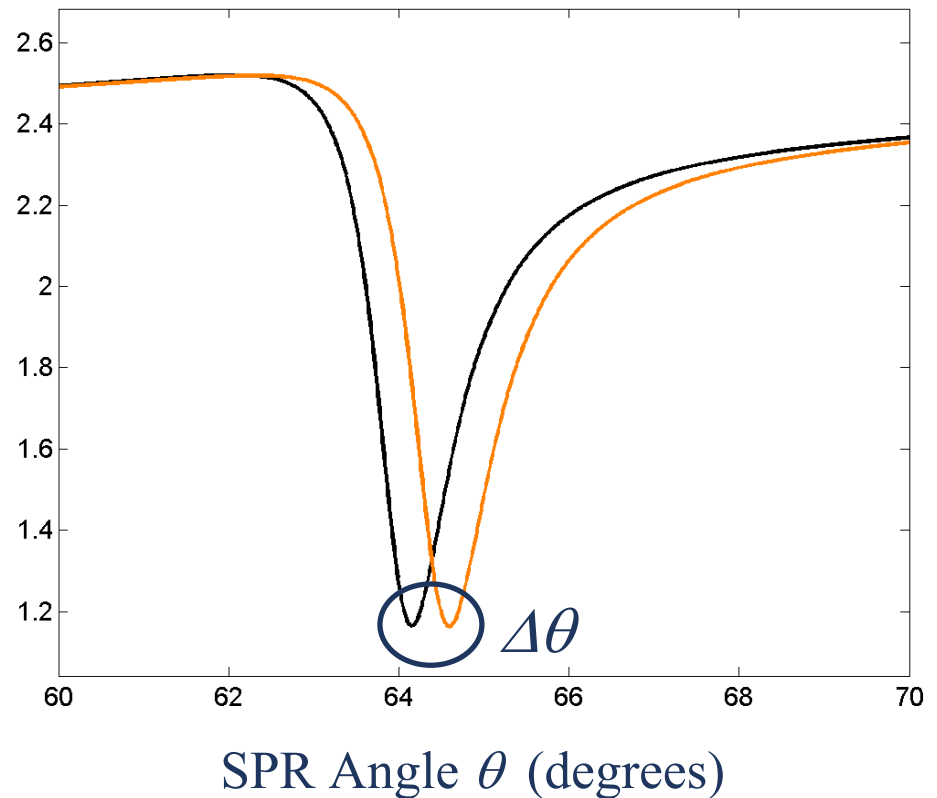
SPR is sensitive to the **thickness d** and the **refractive index n** of dielectric added layers above specific metals (gold, silver, ...).

Buffer
Gold
Prism
Laser

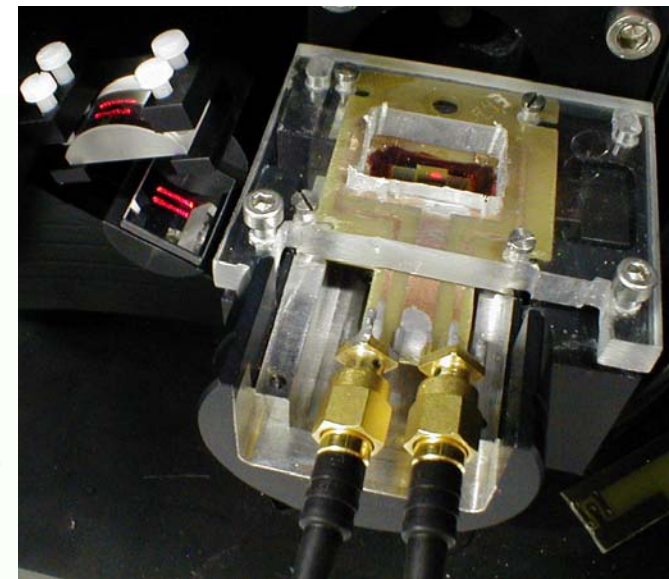
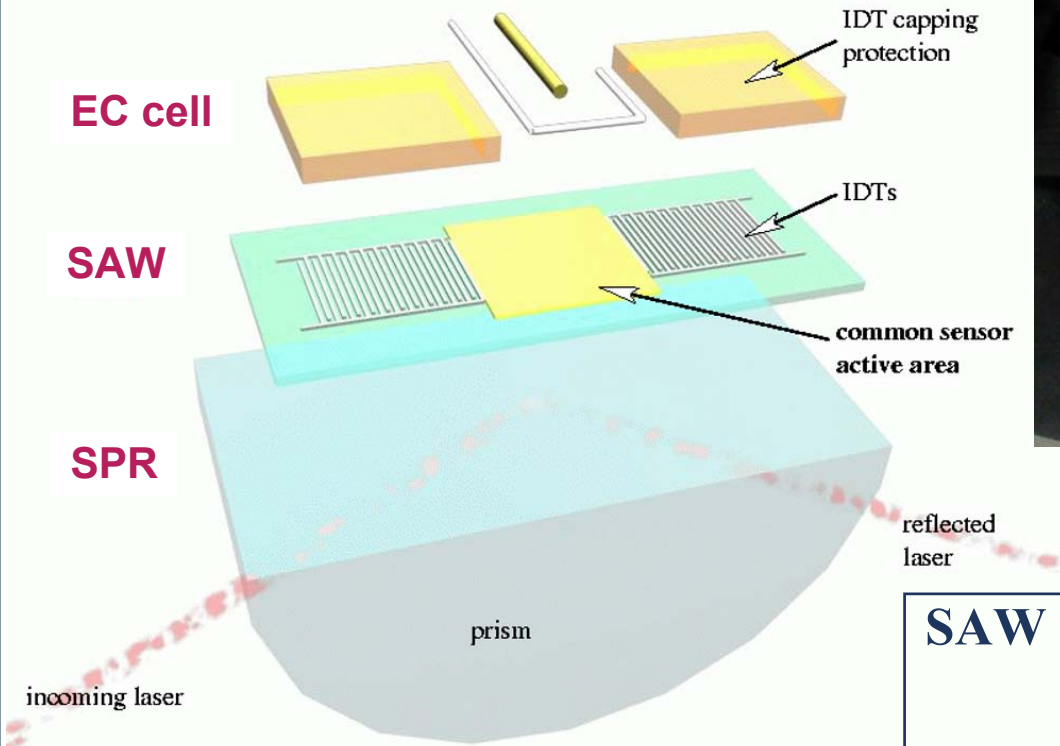


Light intensity
(a.u.)

Simulated SPR signal



COMBINED SAW/SPR: SET-UP



SPR modified IBIS II SPR (IBIS Technologies BV)

670 nm laser light

Kretschmann configuration

SAW ST-cut quartz substrate
1.2 μm H-rich SiO_2
10 nm Ti/ 50 nm Au

wavelength 40 μm

SU8/glass IDT capping

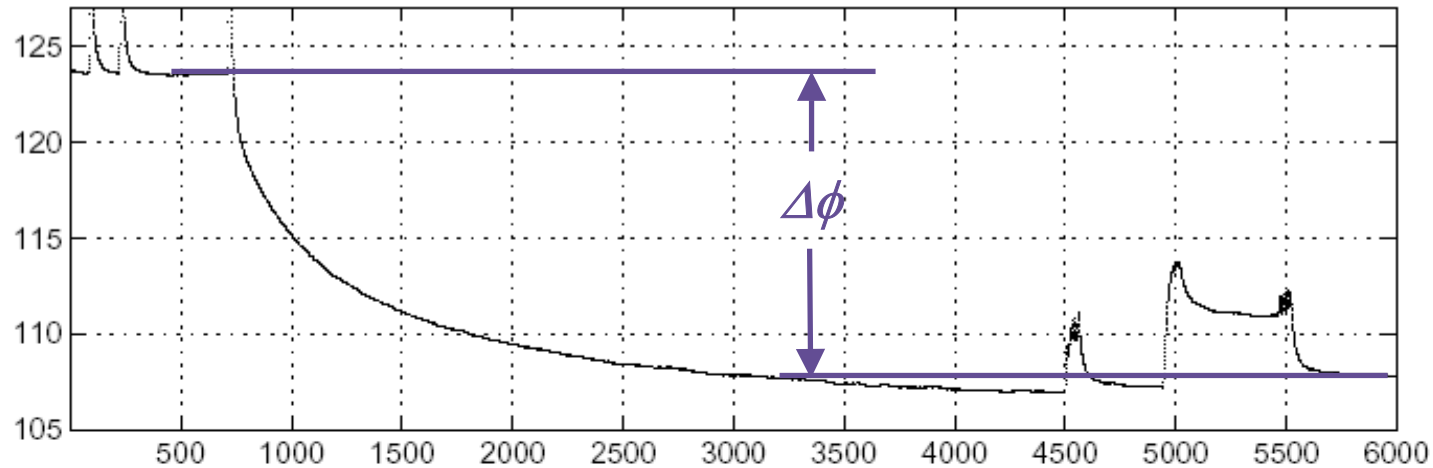
Cu-calibrated mass sensitivity of $-260 \text{ cm}^2/\text{g}$

COMBINED SAW/SPR: EXPERIMENTAL EXAMPLE

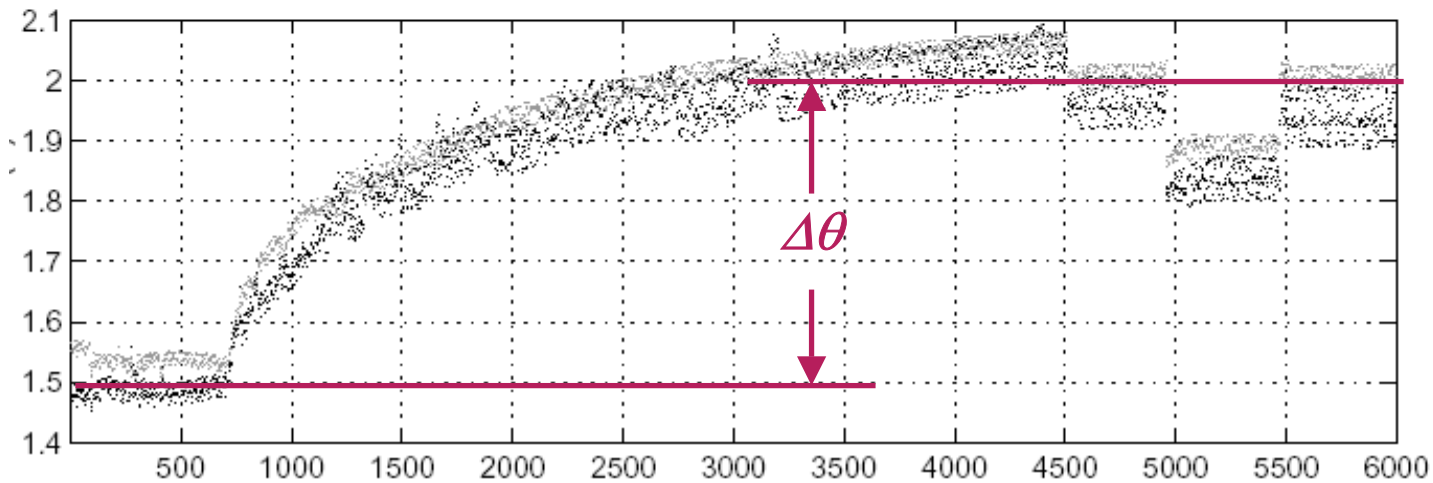
Collagen 30 $\mu\text{g}/\text{ml}$ adsorption on hydrophobic surface



SAW
Phase ϕ

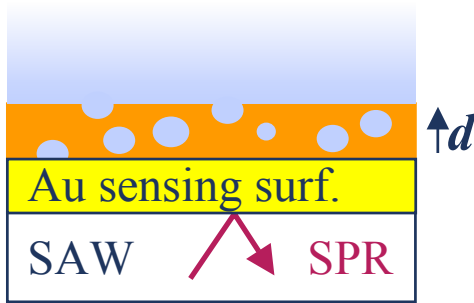


SPR
Angle θ



Time (s)

SAW/SPR MODELING



d = thickness of the layer (protein + water)

x = proportion of proteins in the layer

ρ = density

n = refractive index

Linear combination of the protein and the water:

density

$$\rho(x) = x \rho_{PROTEIN} + (1 - x) \rho_{WATER}$$

refractive index

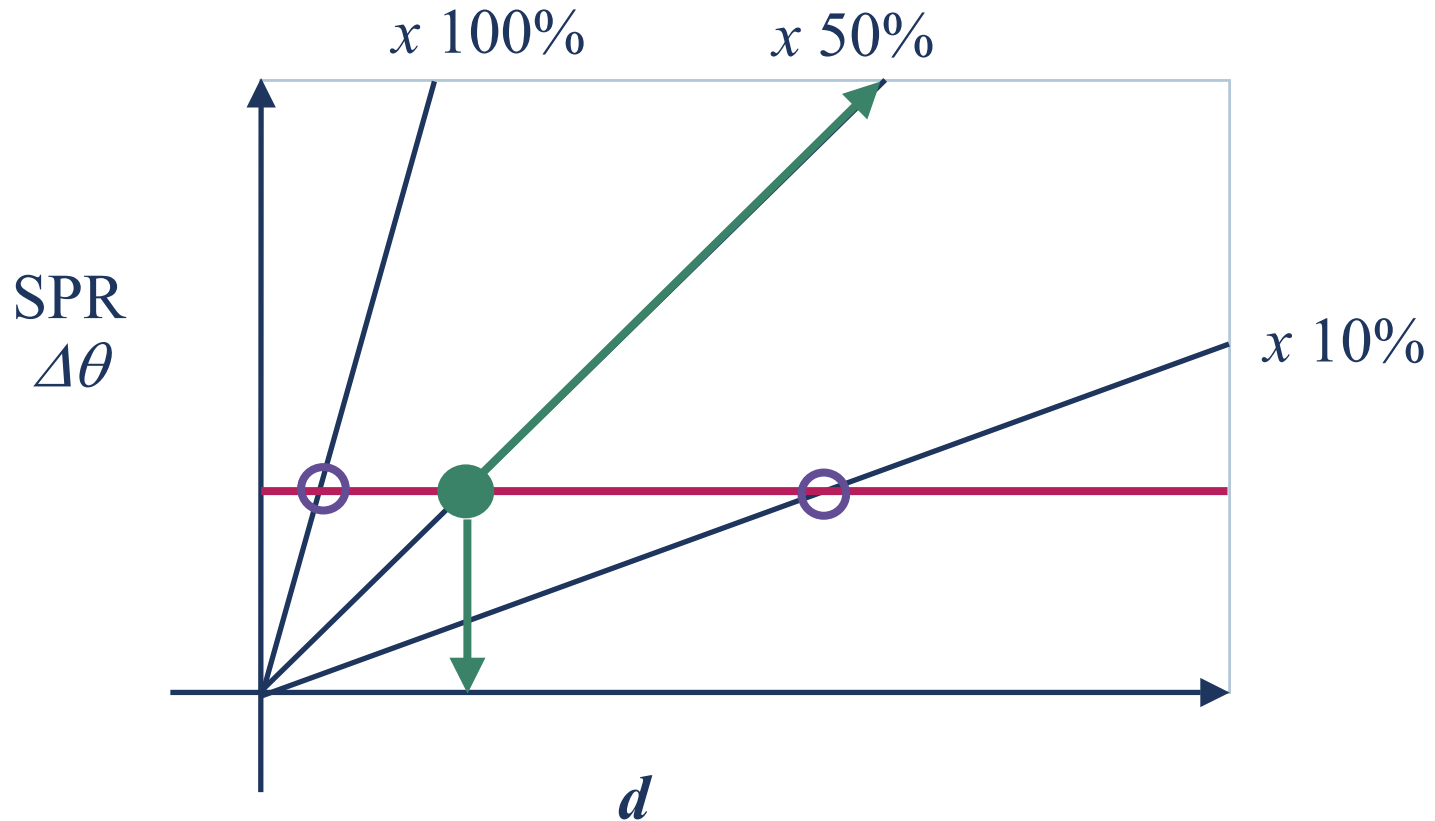
$$n(x) = x n_{PROTEIN} + (1 - x) n_{WATER}$$

The **SAW shift** $\Delta\phi$ gives the surface density m of the layer:

$$m = \left(\frac{\lambda}{360^\circ DS} \right) \Delta\phi = \rho(x) d$$

The **SPR shift** $\Delta\theta$ is simulated from d and $n(x)$.

SAW/SPR MODELING



- 1) Simulated SPR shifts $\Delta\theta(d,x)$
- 2) **Experimental SPR shift**
- 3) Allowed values for SAW shifts $\Delta\phi(d,x)$
- 4) **Extraction of thickness d and protein content %**

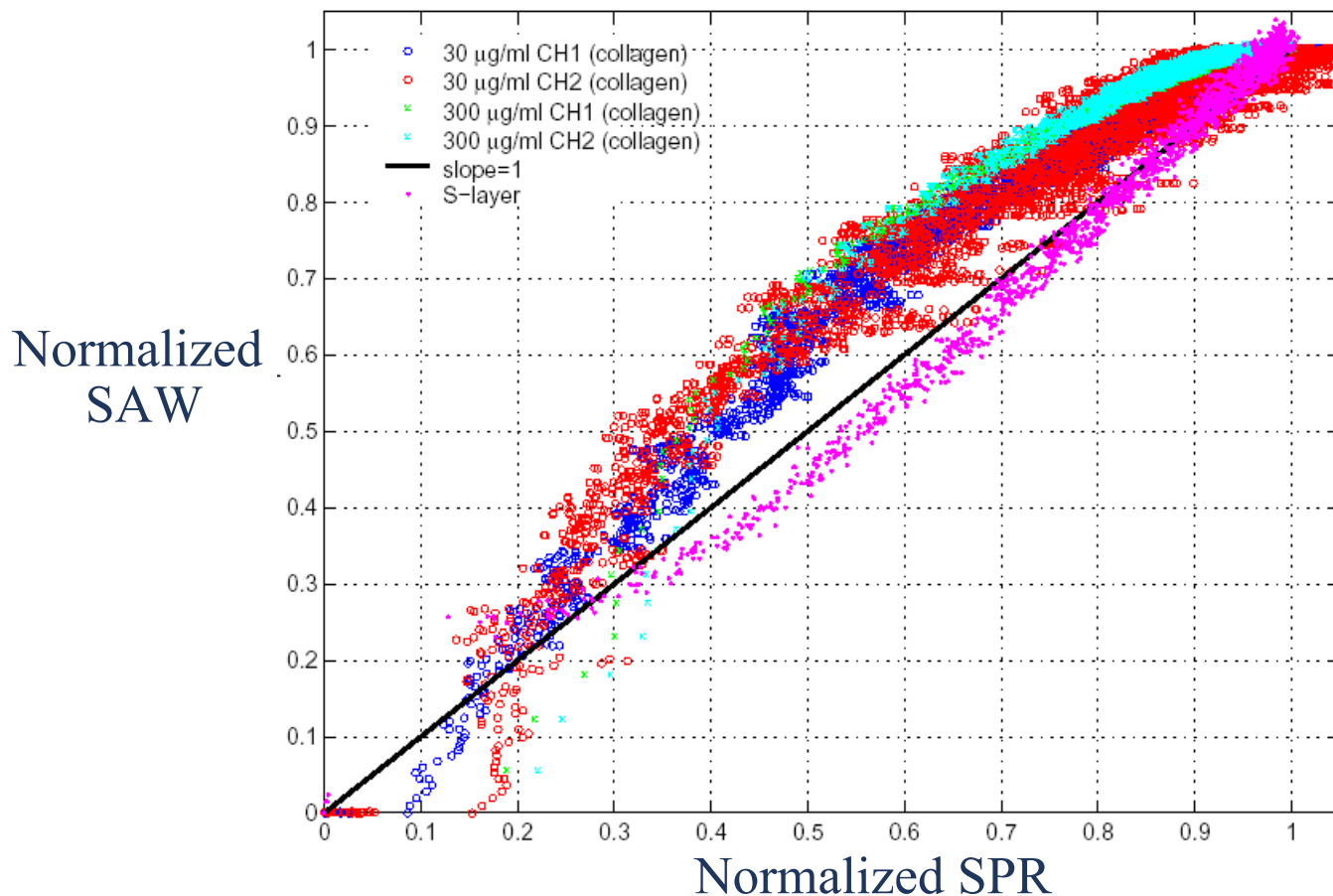
EXPERIMENTAL RESULTS

In-situ measurements of collagen and fibrinogen adsorbed on hydrophobic surfaces:



	Analyte ($\mu\text{g/ml}$)	Surface density m (ng/cm^2)	Thickness d (nm)	Protein content x (%)
STRONG HYDRAT.	Collagen 30 $\mu\text{g/ml}$	1750 ± 150	16 ± 3	25 ± 15
	Collagen 300 $\mu\text{g/ml}$	2100 ± 200	19 ± 3	35 ± 10
STACKED LAYERS	Fibrinogen 46 $\mu\text{g/ml}$	750 ± 100	6 ± 1.5	50 ± 10
	Fibrinogen 460 $\mu\text{g/ml}$	1500 ± 500	13 ± 2	50 ± 10
“DRY” LAYERS FOR SAW CALIB.	S-layer	560 ± 20	4.7 ± 0.7	75 ± 15
	CTAB	135 ± 15	1 ± 0.1	100

EXPERIMENTAL RESULTS: ADSORPTION KINETICS



Normalized
SAW

Normalized SPR

The kinetic of the adsorption monitored by the two techniques is different:

- wrong model ? **Viscous effects** neglected in the SAW !
- mass over/underestimation.

DISCUSSION: DRAWBACKS OF THE TECHNIQUES

SAW drawbacks

- high temperature sensitivity
- rigid mass and viscous effects combined
- signal distortion due to interferences
- different phase and group velocities

SPR drawbacks

- temperature dependence
- signal distortion due to birefringence
- optical interferences for multiple layers
- data extraction through modeling
- limited to metal surfaces (gold, silver)

Combined technique

- no interferences between SAW and SPR
- complementary information allowing extraction of coupled parameters ($\rho, n \rightarrow x, d$) on a same layer

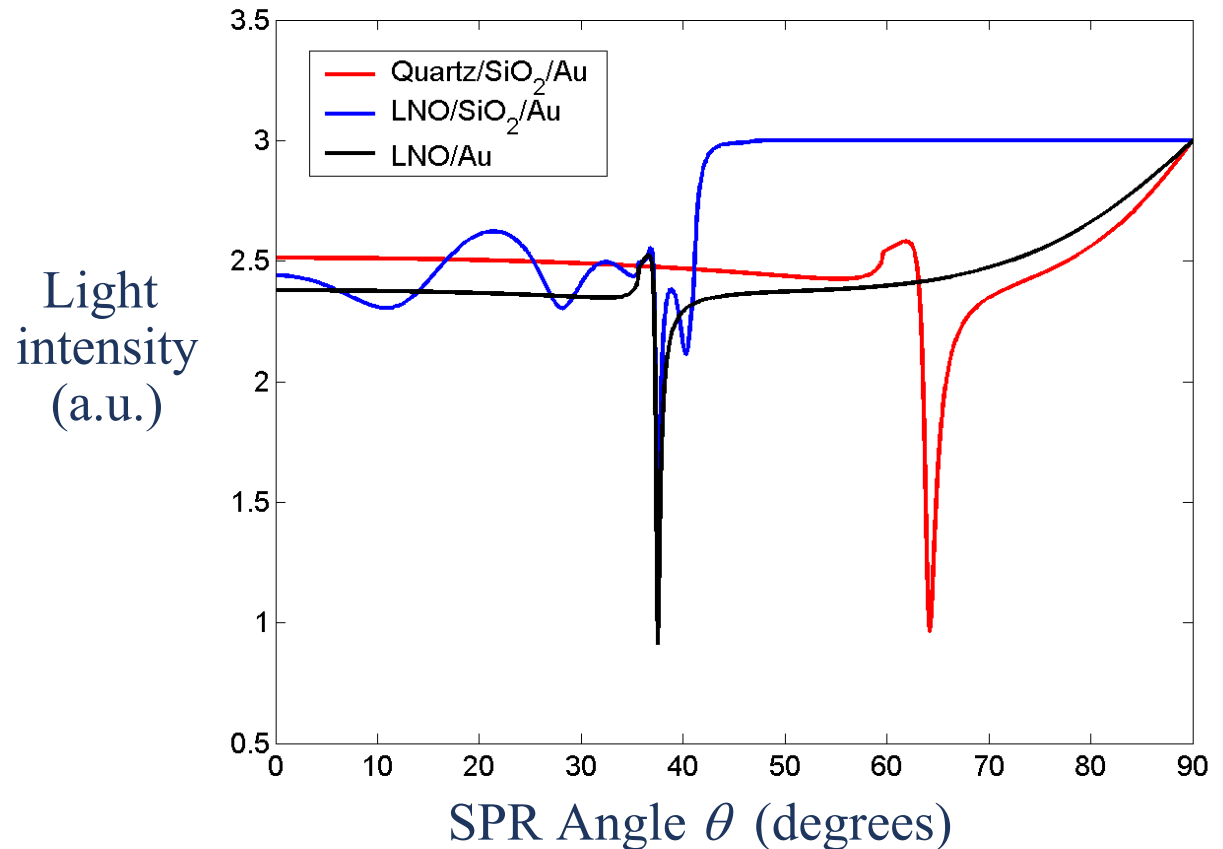
DISCUSSION: LOVE MODE SAW MATERIAL COMBINATIONS

SPR angle is function of n and of the light wavelength.

Interferences and total internal reflection effects must be considered for a stack of layers.

SPR sensitivity higher for θ closer to 90° .

Conductive layers influence the SAW transfer function by shortcircuiting the transducers.



CONCLUSIONS

- Combined SAW/SPR technique:
 - provides information about the **thickness** and **water content** of protein layers;
 - **real time measurements**, with some insights into the adsorption kinetics;
 - **label-free** acoustic and optical method.

- Ways to increase the Love mode SAW mass sensitivity:
 - H-rich PECVD SiO₂
 - Gold

- Experimental results on collagen and fibrinogen adsorption demonstrate the potential and the limits of the combined technique.

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Slides of this presentation available at URL
<http://friedtj.free.fr/chua/biosensors2004.pdf>
or by e-mail : francisl@imec.be